A rapid and highly predictive chemiluminescent cell-based assay to monitor intracellular xanthine oxidase activity for the screening of natural inhibitors in living endothelial cells

Cristiana Caliceti^{*†1}, Donato Calabria¹, Martina Zangheri², Massimo Guardigli¹, Mara Mirasoli¹, Patrizia Simoni³, and Aldo Roda¹

¹Department of Chemistry "Giacomo Ciamician" and "Centro Interdipartimentale di Ricerca Industriale Energia e Ambiente" Alma Mater Studiorum, Università di Bologna, Bologna, Italy – Italy ²Department of Chemistry "Giacomo Ciamician", Alma Mater Studiorum, Università di Bologna, Bologna, Italy – Italy

³Department of Medical and Surgical Sciences (DIMEC), University of Bologna, Alma Mater Studiorum, Sant'Orsola Malpighi Hospital, Bologna, Italy. – Italy

Abstract

Xanthine oxidase (XO), an enzyme expressed at high levels in vascular endothelium, catalvses the oxidation of hypoxanthine to uric acid, linked to gout and cardiovascular diseases. Testing the inhibition of XO is crucial to identify potentially effective natural products that could be used to treat several diseases. Herein, we report the development of a fast, reliable and highly predictive chemiluminescent (CL) bioassay to determine XO activity into living endothelial cells and its use to obtain quantitative information on the XO inhibiting activity of natural products of great interest in the nutraceutical field. The method is based on the intracellular CL emission of luminol, caused by XO-derived H2O2, employing Fe2+-EDTA complex as catalyst in 0.1M borate buffer (pH=10.3). The assay, which could be completed in less than 30 minutes and displayed a limit of detection of $0.4\mu U/mL$, allowed measuring intracellular XO activity, which was assessed to be $(1.3\pm0.3)\times10$ -7mU/cell in human vascular endothelial cells (HUVEC). Next, to demonstrate the applicability of the CL cell-based assay to screen potential XO inhibitors, HUVEC cells were treated with different natural extracts, utilizing oxypurinol, the active metabolite of the drug allopurinol, as standard (IC50 152±76 ng/mL). The capability of several natural products to inhibit XO activity was evaluated, obtaining an IC50 of $28\pm4\mu g/mL$ and $14\pm3\mu g/mL$ for Ganoderma Lucidum and Cordyceps Sinesis fungi, respectively; $0.86\pm0.07\mu$ g/mL and $1.27\pm0.08\mu$ g/mL for ultrasonicassisted extracted (UAE) and Naviglio®-assisted extracted (NAV) winemaking by-products, respectively; 120 ± 50 mg/mL and 150 ± 40 mg/mL for yeast fermented papaya (FPPA) and Lactic Acid Bacteria fermented papaya (FPPB), respectively. Since the bioavailability of the compounds, especially the ability to cross cell plasma membranes, is an important issue to take into consideration for nutraceutical applications, this method, which employs whole cells, is highly representative and predictive to study intracellualr XO inhibitors in vitro.

 $^{^*}Speaker$

 $^{^{\}dagger}$ Corresponding author: cristiana.caliceti@unibo.it

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